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Development of a SIMS compatible internal standard spiked resin for quantitative bio-imaging of biological samples by laser ablation ICP-MS: an application to uranium contaminated kidney samples

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Context

It would be desirable that both samples and calibrations standards could be shared by different imaging techniques. The aim of this work is to develop a sample/standard preparation methodology compatible with laser ablation and Secondary-Ion Mass Spectrometry (SIMS) analyses for a complementary analysis of uranium distribution in kidney with both techniques. On the other hand, a novel approach based on internal standard doped resin was carried out in order to incorporate a more feasible IS than ¹³C (1). Therefore, thulium spiked pure EPON resin was synthetized and employed to embed dehydrated samples and doped kidney homogenates based on the chemical sample preparation protocol of biological samples for SIMS analysis (2, 3). Nevertheless, the chemical dehydration protocol was adapted to the viscous and liquid state of the homogenates. In parallel, in order to replace the use of organ homogenates for calibration standards (laborious task and required access to animal facilities), uranium spiked resin was also tested. Then, serial thin and ultra-thin sections were cut with microtome and the homogeneity of internal standard was evaluated by randomly analysing small selected areas by LA-ICP-MS and SIMS. Laser ablation conditions were optimized to achieve complete sample consumption of the tissue whilst minimizing the penetration into the glass slide.

Experimental procedure



Conclusions

We developed an analytical methodology based on internal standard spiked resin which will allow the quantification by LA-ICP-MS the resin embedded samples used for other imaging techniques like SIMS microscopy. The advantages of this new methodology are 1) the addition of a suitable internal standard to both matrix-matched standards and biological samples without altering their original uranium distribution, 2) an appropriate sample preparation compatible with several imaging techniques (SIMS, TEM, LA), 3) ease to prepare and 4) room temperature storage as a solid material which would facilitate its transport. Some optimisations are still needed, in particular to control the resin thickness. This methodology will positively contribute to the collaboration among bio-imaging techniques users.



Acknowledgments

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