

# Minimum Reporting Standards Should be Expected for Preclinical Radiobiology Irradiators and Dosimetry in the Published Literature

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1	Commentary: Minimum Reporting Standards Should be Expected for Preclinical Radiobiology
2	Irradiators and Dosimetry in the Published Literature
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### Introduction

Preclinical researchers have long acknowledged the critical need for accurate dosimetry in the conduct of radiation-exposure studies involving living systems. Although radiation dosimetry is required to be National Institute for Standards and Technology (NIST)-traceable for human irradiation protocols, there were previously only limited requirements stipulating accurate radiation exposures for preclinical radiation studies funded by the National Institutes of Health (NIH). In one of the earliest identified publications (1956) that addressed radiation dosimetry in murine studies, Hurst *et al.* (1) discussed their calculations surrounding estimating neutron responses, and "distribution of of absorbed energy versus LET". In 2009, Zoetelief *et al.* (2) outlined European efforts to harmonize X-ray dosimetry studies in radiation biology, which resulted in the establishment of a protocol to improve dosimetry oversight and allow for better comparison of results across different institutions. It is, however, unclear if these early efforts substantially changed the approach of biologists to ensuring accurate and reproducible dosimetry in their radiation research.

The lack of consideration of dosimetry in *in vitro* and *in vivo* work was further explored in 2011 during the "Radiation Dose is More than a Number" workshop convened by NIST, and co-sponsored by the National Institute of Allergy and Infectious Diseases (NIAID), and the National Cancer Institute (NCI). The report that followed (3) made nine recommendations that ranged from coordinating with radiation physicists during the experimental design phase and including more setup and dosimetry details in publications, to establishing working groups to develop protocols, and implementing formal dosimetry intercomparison programs. In an attempt to address the latter suggestion, and ensure rigor and reproducibility in their funded research portfolio consistent with NIH grants policy<sup>1</sup>, the NIAID Radiation and Nuclear Countermeasures Program (RNCP) released a request for proposals (NIAID-NIH-RFP-NIHAI201800020<sup>2</sup>) in 2019, seeking to make a single contract award for "RNCP-Wide Dosimetry Guidance & Monitoring of Sources and Irradiation Protocols". This funding opportunity requested that offerors develop a consistent means of dosimetry comparison and reproducibility, as well as provide the administrative foundation necessary to

<sup>&</sup>lt;sup>1</sup> https://grants.nih.gov/policy/reproducibility/index.htm

<sup>&</sup>lt;sup>2</sup> https://sam.gov/opp/360d070bdf5d437f83e88c13442fa23f/view

facilitate and coordinate dosimetry activities in partnership with the NIAID. Respondents were asked to provide services, facilities, expertise, and capabilities to develop a centralized dosimetry harmonization effort that fits the resources and circumstances of projects across the funded RNCP portfolio. The contract award was made in 2020 to the University of Wisconsin (Principal Investigator Larry DeWerd), who established a harmonization protocol to encompass all the irradiators in use across the RNCP-funded portfolio of grants, contracts, and inter-agency agreements.

In 2019, the RNCP also became concerned with the specifics of establishing and reporting neutron dosimetry and convened a "Neutron Radiobiology and Dosimetry Workshop" with presentations from 17 subject matter experts, planned in collaboration with the Department of Defense (DoD), Defense Threat Reduction Agency (DTRA), and the National Aeronautics and Space Administration (NASA). The meeting report that was published in 2021 (4) explored historical neutron exposure research and highlighted the need for harmonized reporting of exposure and experimental parameters. The authors concluded that efforts should be made to update computer coding for neutron dosimetry estimates, government policies needed to be re-visited in terms of expected radiation energy spectrums, and funding agencies must make key investments in consideration of neutron dosimetry, to improve science and ensure radiation emergency preparedness.

All of these early attempts to ensure the use of accurate dosimetry in research laboratories culminated in the current effort to take the next step and make strong recommendations that sufficient information be included in manuscripts or supplemental data files, such that reviewers and editors can be assured that the irradiations were carried out correctly, and so readers of the published work have the necessary information to replicate published experiments.

## **Background and Rationale for Dosimetry Reporting**

Good science must be well described to be understood, compared, and replicated. For radiation biology, this requires the combination of both biology and radiation physics – two very different expertises, typically provided by different individuals with specialized training. While biological systems can be

difficult to control, particularly for *in vivo* models, radiation follows well-understood physical processes, which can be controlled and measured to a very high level of accuracy, precision, and reproducibility. Therefore, it is both feasible, and important, to require radiobiological studies to describe their irradiation protocol.

Unfortunately, experimental irradiation protocols are currently not well described in the majority of published radiobiology studies. This was already true in 2011, during the original NIST workshop (3), and has since been confirmed by multiple meta-studies showing no apparent improvements in reporting since the original symposium (5-9). In a recent analysis of 1758 published pre-clincal studies employing ionizing radiation, some parameters, such as radiation dose, are nearly always provided (>94%). Many important details, such as the equipment producing the radiation, the radiation type, beam energy (or quality), and dose rate, are usually, but not always, reported (>50-80%). Details pertaining to experimental design, such as the number and geometry of the radiation fields, the use of shaping collimators, or rotating tables to ensure uniform delivery to multiple animals, are seldom shared (>25%). Finally, details relating to experimental validation of the delivered radiation dose through some form of dosimeter (ideally traceable to national standards), are nearly always absent (<15%). Reference to published standards or protocols was as low as 1.3% of published studies in Draeger *et al.* (9).

As originally illustrated in the report following the NIST workshop, "investigators cannot report what they have not considered." (3). When experiments are not described, a reviewer cannot assess what an author might or might not have considered, or the soundness of the experimental design at all. In some instances, metastudies reveal that certain publications contained glaring contradictions in their irradiation protocols. These include descriptions of irradiators that would be incapable of producing the experiment described in the methods, non-existent radionuclides, or typographical errors (e.g., kV to MV), which translate into a thousand-fold difference in energy. These findings suggest that many studies may be erroneously described, even in cases where there is no clear contradiction in the few details available. For instance, irradiating multiple mice placed at varying distances from the center of the field will often result in

a different dose (and therefore a different biological effect) delivered to each, but this will not be apparent in a manuscript that simply describes the source used and dose prescribed.

In a vacuum, a lack of rigor in describing experimental protocols might not imply anything, but insufficient communication among authors, or a propensity to copy and combine old protocols without sufficient rigor can be problematic. When combined with repeated reports outlining difficulties in harmonizing radiation dosimetry across separate laboratories, (2, 7, 8) a more concerning picture emerges. As stated in the original NIST report, "Few students or researchers using ionizing radiation in biological research have training in basic radiation physics. This leads to the difficult situation that when 'one does not know what one does not know' dosimetry design and documentation go unaddressed". And if it is not described, a journal or grant reviewer cannot assess whether the experiment is sound.

One particular concept not initially addressed in the first NIST workshop concerns the definition of radiation dose. Indeed – not all dose is created equal. An X-ray tube can be calibrated in terms of exposure, air kerma, or adsorbed dose to water  $(D_W)$ , depending on which correction factors are used. Reference dosimetry can assume, or not, the presence of backscatter. Or, an investigator can refer to the radiation dose being delivered to animals under experimental conditions. Each of these descriptions is scientifically sound, and yet can lead to differences of up to 50% in reported doses. As described in the next section, the use of Roentgen (exposure) vs dose for  $^{137}$ Cs exposures will introduce an error of  $\sim$ 11%. Without further description, it is impossible to know which quantity an investigator is referring to. To address this, the American Association of Physicists in Medicine (AAPM)'s Task Group 319 is developing new standards on how to perform and report experimental dosimetry in kilovoltage cabinet irradiators. However, these new guidelines have yet to be published, and their implementation will not be instantaneous. In the meantime, investigators should be careful in how they describe their irradiation protocol in scientific communications.

#### **U.S. Efforts in Dosimetry Harmonization**

Inconsistent dosimetry was first outlined by the 2011 NIST panel<sup>3</sup>, and follow-up publications in 2016 by Seed, *et al*, (8) and Pedersen, *et al*. (7) suggested that only one in five laboratories deliver dose within 5% of a specified target level, with variations up to 42% observed. From 1971 to 1999, the EULEP-EURADOS initiative performed 8 similar intercomparisons and consistently found errors of up to ~20%, and found the majority of institutions could not satisfy a  $\leq$ 10% homogeneity requirement (*10*). Radiation biology research often employs orthovoltage X-ray systems and <sup>137</sup>Cs sources where dosimetry is based on manufacturer recommendations. For X-rays, results may not be traceable to NIST standard beams because of the differing maximum beam energy and the filtration used for a certain energy. In addition to traceability issues, low-energy X-rays introduce variability in dosimetry and radiobiological response in multiple ways. First, low energy bremsstrahlung radiation is more readily absorbed in tissues than are higher energy photons from isotope irradiators. Second, low-energy X-rays are more readily absorbed by cortical bone, inducing differential biological effects in bone marrow and different biological responses (*11-13*).

From 2020 to 2023, The NIAID/RNCP program for harmonization of radiation dose evaluated >30 irradiators (X-ray and cesium), and found that upon initial survey, 50% of machines were outside of  $\pm$  5% of the targeted 4 Gy dose, the benchmark for acceptable irradiation, as is shown in Figure 1. Traceable NIST beams for X-ray beams were used for cabinet X-ray systems by matching half value layers (HVL) for radiobiological X-ray beams, although kVp and second HVL are much different. Phantoms for total-body, and partial-body irradiation were established, and dose was measured using LiF:Mg,Ti thermoluminescent dosimeters (TLDs) calibrated using the NIST-traceable HVL matched X-ray beams.

Variations from the 4 Gy target dose ranged from +/-12% to 25%. After consultations and retesting, results for nearly all institutions were brought within the required 5% of target dose, and doses were resolved for the remaining institutions after consideration of physics issues. Also, precision mapping of the radiation field using multiple TLDs was important and permitted the resolution of dose discrepancies that aligned with variations in dose. To summarize, there are three problems that are frequently encountered at institutions that must be addressed. First, the correct physics dose quantity and other characterizations

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<sup>&</sup>lt;sup>3</sup> https://www.nist.gov/programs-projects/medical-archive-workshop-radiobiology-dosimetry-standardization.

should be employed, especially for X-ray machines. An example of this that was found with some radionuclide sources involving a dose discrepancy of ~11 %. The institutions in question were using the manufacturer-supplied radiation exposure quantity in Roentgens, rather than the absorbed dose to water; and the correction needed to achieve absorbed dose to water was 11% - matching the observed error perfectly. A second problem is not understanding the physics or the measurements given; consultation with a medical physicist would be of great benefit, but not all sites conducting radiation biology research have access to the support of a certified medical physicist. Of note, not all certified medical physicists are familiar with irradiation conditions and calibration protocols of preclinical systems, which can be vastly different than clinical systems. Finally, an inadequate understanding of dose distribution within the radiation field, especially in cabinet-style orthovoltage X-ray irradiators, is a significant problem. It is important to understand the isodose plot of the field, and in many cases, rotation of the subject animals is necessary to average out the dose. The goal of this NIAID-funded program is the harmonization of dose among institutions regardless of the source used to establish the radiation.

## **French Efforts in Dosimetry Harmonization**

In France, the Institute of Radiological Protection and Nuclear Safety (IRSN) has a long tradition of collaboration between dosimetrists and radiobiology researchers. Technical support has always been provided to define the best irradiation conditions and reference dose rates, whether for gamma irradiators, conventional X-ray facilities, or for irradiation at reactors (e.g., the SILENE facility in Valduc, France) with a concern for the traceability of these data. A dose rate in the most appropriate dosimetric quantity and a homogeneity level are defined for each irradiation configuration. For example, for cell irradiations, a configuration is defined for a given container, filling volume, and number of containers. It is not only a question of defining a reference rate in D<sub>W</sub> or in air kerma according to the reference protocols in use (such as the AAPM's TG-61 or the IAEA TRS-398), but also taking into account factors that can affect the dose rate, and defining it at the level of the biological material of interest (14). For small animals, whether for total-body or localized irradiations, specific phantoms are produced using molds. For programs involving

study of bone tissue, electron paramagnetic resonance (EPR) spectroscopy, which enables the dose actually delivered to the bone to be estimated, has been introduced (15). These skills have been regularly made available to other national or European research institutes, private companies, and European projects such as RENEB (16). In Figure 2, the accuracy of the reference dosimetry of beams use to establish calibration curves in biological dosimetry was evaluated with alanine dosimeters calibrated in Dw. Deviations from reference dose (10 Gy) range from a few % up to 50%, with about 50% of results within +/- 5%. Dose quantities used, when known by participants, were either air kerma or Dw. The impact of using different dose quantities and/or beam quality was noticeable in dicentric assays, but had not been accounted for when analysing results of biological dosimetry for this inter-laboratory comparison. With the arrival of more complex new small animal X-ray conformal irradiation platforms to the IRSN (SARRP, Xstrahl), and the increased use of a linear accelerator (LINAC), the need to have a dedicated physicist on a full-time basis was felt to be important, due to the greater complexity of use and dosimetry aspects. This recommendation has also resulted in a dedicated research program at IRSN on the latter aspect, notably to develop new dosimetry approaches (e.g., for small fields), new protocols for dose rate measurement at biological sites, verification of manufacturer data, and quality control. A similar need among other users of SARRP type of X-ray platforms has resulted in hiring additional physicists to ensure the optimal performance of these machines. This small community quickly organized itself to share experiences, protocols, and methods, and endeavored to set up a program to verify manufacturer data and provide intercomparison and technical assistance for machines without local physicist expertise, notably in the framework of a new French national network of irradiation platforms for radiobiology studies (Réseau des Plateformes National d'Irradiation; ResPlanDir)<sup>4</sup>.

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## **Proposed New Instructions for Authors for Dosimetry Reporting**

Given the concerns outlined above regarding accuracy and reproducibility of radiation dosimetry in radiobiological research, and in the interest in ensuring rigor and reproducibility of irradiation experiments,

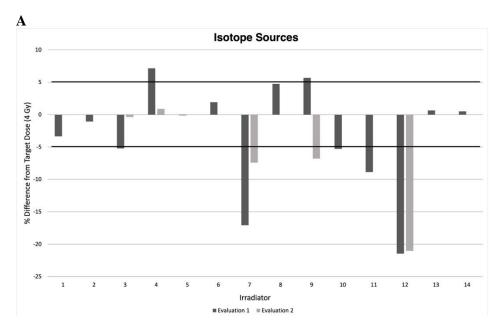
<sup>&</sup>lt;sup>4</sup> https://www.france-hadron.fr/en/plateforms/features-modalities.html

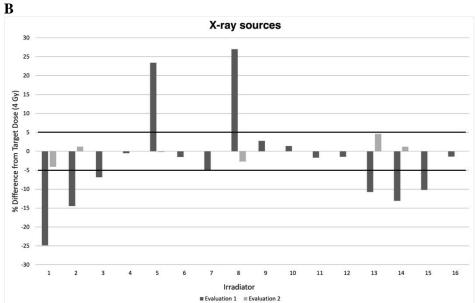
the authors of this commentary, in agreement with another recent commentary on this subject by Marcu e
al. (17), propose that journals engaged in publishing radiobiology research introduce new 'Instructions for
Authors' addressing minimum standards for reporting radiation dosimetry information. Implementing the
recommendations below would also address issues sometimes encountered in successful study replication
Therefore, the following minimum irradiation parameters are suggested:

- Name of the irradiator (company, country, model)
- Type of source (radionuclide (e.g. cesium, cobalt), X-ray)
- Energy (kVp, mA; for X-ray systems)
  - Filtration (for kV X-ray systems, half value layer if known)
  - Radiation dose and dose rate, source to surface distance (SSD), field size,
- Sample being irradiated, and

- Irradiation geometry (nature of animal restraint (e.g., pie or other jig), the material of the sample holder, shielding (if appropriate), orientation to the source, any animal(s)/sample rotation)
- Dose quantity (e.g., how radiation dose was measured D<sub>W</sub>, air kerma, midline tissue, etc.)
- Daily pre-exposure device calibration method
- Planning and dose calculation method (required for multiple field conformal dose delivery,
  particularly orthovoltage small animal conformal dose delivery systems, optional otherwise)
- Reporting of any in-run dose measurements (if performed)

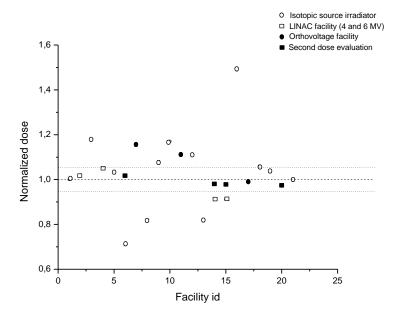
## Figure 1





**Figure 1.** A) Performance of U.S. institutions with isotope irradiators within NIAID-supported sites; B) Performance of institutions with orthovoltage X-ray irradiators within NIAID-supported sites. The bold horizontal lines in each panel represent the  $\pm$ -5% cutoffs for acceptable variance from the 4 Gy target dose. Gray bars indicate paired final retesting results for irradiators with initial test doses that were  $\geq \pm$ -5% of the target dose.

## Figure 2



**Figure 2**: Doses measured at 21 European facilities with alanine with a calibration in terms of dose in water and normalized to the dose delivered at the French primary laboratory (modified and used with permission from) (16)).

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